

Study of the Role of Antioxidant (Vitamin C) on Modulation Toxicity of Chronic Use of Monosodium Glutamate in Liver of Albino Rats

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Abstract

Introduction: Monosodium glutamate (MSG) is now used in many foodstuffs as a food additive and flavour enhancer. Although it has classified as safe food ingredient, the use of MSG remains controversial. MSG is a slow excitotoxin food additive which can cause generation of numerous amounts of free radicals which affects many organs such as liver. Vitamin C is an antioxidant and inhibits chemical carcinogenesis by protecting the body against oxidative stress. **Aim of the study:** Study role of antioxidant (vitamin C) on modulation biochemical, histological, histochemical and ultrastructural changes of liver caused by chronic use of MSG. **Materials and Methods:** Sixty adult albino rats divided into equal three groups, first group (control) received 1ml of saline daily for three months, Second group received MSG 6mg/g/BW/day for three months, third group received MSG 6mg/g/BW/day and vitamin C 500 mg /kg /b.w /day, orally and for three months. Biochemical changes were investigated by the liver function tests. Assessment of histopathological changes of liver was done by using light microscope, transmission electron microscope, histochemical studies and immuohistochemical studies. **Results:** After chronic use of MSG, light microscope and transmission electron microscope examination revealed severe histopathological changes such as hepatic architecture destruction and dilatations of the central veins with statistical significant increase of liver enzymes. Histochemical studies revealed severe reduction of carbohydrates and proteins; pyknotic nucleus, vacuolated cytoplasm, swollen mitochondria and vesiculated rough endoplasmic reticulum with significant positive stained nuclei with ki-67 and p53. Administration of Vitamin C with MSG led to significant improvement of biochemical and pathological changes of liver. **Conclusions:** Chronic use of MSG caused hepatotoxicity of rats which is improved by administration of Vitamin C with it.

Keywords Monosodium glutamate, Vitamin C, Liver

Introduction

Monosodium glutamate (MSG) is used commercially as a food additive and is commonly marketed as a flavour enhancer. It is now used by most fast-food chains and in many foodstuffs, in particular, processed foods (Leung and Foster, 2003).

Trade names of monosodium glutamate include Ajinomoto, Vetsin, Ac'cent and Tasting Powder (Beyreuther et al., 2007). It was once made predominantly from wheat gluten, but is now made mostly from the fermentation of carbohydrates with a nitrogen source, using bacterial or yeast species from

genera such as *Brevibacterium*, *Arthrobacter*, *Microbacterium*, *Micrococcus* and *Corynebacterium* (Willams and Woessner, 2009).

MSG is a slow poison which hides behind dozens of names, such as natural flavouring and yeast extract. It is not a nutrient, vitamin, or mineral and has no health benefits. The part of MSG that negatively affects the human body is the "glutamate", not the sodium. The breakdown of MSG typically consists of 78% glutamate, 12% sodium, and about 10% water (Stevenson, 2000).

Any glutamate added to a processed food is not considered naturally occurring. Natural glutamate in plants and animals is known as L-glutamic acid. Our normal digestive process slowly breaks down this natural or "bound" glutamic acid and it is then delivered to glutamate receptors in our body and brain. Broken down this way, it is harmless. In a factory, however, the bound glutamic acid in certain foods (corn, molasses, wheat) is broken down or made "free" by various processes (hydrolyzed, autolyzed, modified or fermented with strong chemicals, bacteria, or enzymes) and refined to a white crystal that resembles sugar (Hazar et al., 2008).

Although the Food and Drug Administration (FDA) has classified MSG as a food ingredient that's "generally recognized as safe", the use of MSG remains controversial. For this reason, when MSG is added to food, the FDA requires that it be listed on the label (Ortiz et al., 2006).

MSG is toxic to humans and that over 25% of the population suffer adverse reactions from MSG7-36. In general, the natural glutamic acid found in food does not cause problems, but the synthetic free glutamic acid formed during industrial processing is a toxin. In addition, when MSG is formed using hydrochloric acid the final product includes carcinogens (Geha et al., 1998).

MSG is excitotoxin food additives. It is also known that excitotoxins themselves can cause the generation of numerous amounts of free radicals and that during the process of lipid peroxidation (oxidation of membrane fats) a substance is produced called 4-hydroxynonenal. This chemical inhibits the glutamate transporter, thus allowing glutamate to accumulate in the brain. Excitotoxins destroy neurons partly by stimulating the generation of large numbers of free radicals. Recently, it has been shown that this occurs not only within the brain, but also within other tissues and organs as well (liver and red blood cells) (Freeman, 2006).

Effects of excitotoxin food additives generally are not dramatic. Some individuals may be especially sensitive and develop severe symptoms and even sudden death from cardiac irritability, but in most instances the effects are subtle and develop over a long period of time (Kondoh and Torii, 2008).

The liver is the second largest organ in the human body after the lung and it is one of the five vital organs. The liver conducts several hundreds of functions

every second; it metabolizes nutrients and substances, helps with food digestion and cleans the blood. It also stores many vitamins and minerals. The condition of the liver is often ignored until something is discovered to be wrong with the liver. The most common "condition" to affect the liver is toxicity, as defined in the Taber's Medical Dictionary as "being poisoned" (Collison et al., 2009). Rat liver and rat tissues metabolize glutamate by oxidative deamination (or transamination to oxaloacetic or pyruvic acid via alpha-ketoglutarate to succinate (Beyreuther et al., 2007).

Some studies revealed that during intestinal absorption, a large amount of glutamic acid is transaminated and consequently alanine levels in portal blood are elevated. If large amounts of glutamate are ingested, portal glutamate levels will increase, this elevation results in an increase hepatic metabolism of glutamate, leading to release of glucose, lactate, glutamine, and other amino acids, into systemic circulation (Eweka et al., 2011). Another studies found that MSG caused an enlargement of the liver and an increase of serum albumin and decrease in serum globulin (Diniz et al., 2004).

Vitamin C (L-ascorbic acid) is an essential nutrient for humans and certain other animal species. In living organisms, ascorbate acts as an antioxidant by protecting the body against oxidative stress. It is a powerful reducing agent capable of rapidly scavenging a number of reactive oxygen species (ROS). Ascorbic acid is a well known for its antioxidant activity, acting as a reducing agent to reverse oxidation in liquids. When there are more free radicals (reactive oxygen species "ROS") in the human body than antioxidants, the condition is called oxidative stress. It prevents oxidative damage to the important biological macromolecules, such as DNA, proteins and lipids. Some investigators reported that antioxidants inhibit chemical carcinogenesis when the antioxidants are administered either prior or with carcinogen (Padayatty et al., 2003).

Vitamin C exerts protective role against acute ultraviolet B-rays (Sunburn cell formation), organophosphorous pesticides and could reduce aflatoxin induced liver cancer (Meves et al., 2002). Moreover vitamin C abolishes chromosome damage resulted from the effect of toxic substances and help to protect the body against pollutants. Because vitamin C is a biological reducing agent, it is also linked to preventive of degenerative diseases such as cataracts, certain cancer and cardiovascular disease (Barros et al., 2004).

Increased vitamin C intake could possibly reduce and prevent nephrotoxic effect. It assists in the prevention of blood clotting and bruising. It strengthens the walls of the capillaries and it is also needed for healthy gum. Vitamin C helps to reduce cholesterol levels, high blood pressure and preventing atherosclerosis. It protects susceptible cells from genotoxicity associated with antiestrogen metabolite-4-

hydroxyl tamoxifen (4-OH tom) and inhibit DNA adduct induced by tamoxifen (Pavlovic et al., 2009).

There were many previous studies about modulation of MSG toxicity by other materials. Oscar et al., (2006) studied positive effect of vitamin E on monosodium glutamate induced oxidative stress in rats. Hazar et al., (2008) showed hepatoprotective effect of N-acetyl cysteine and carotene on monosodium glutamate induced toxicity. Pavlovic et al., (2009) used the ascorbic acid to modulate monosodium glutamate induced cytotoxicity in rat thymus.

The present study aims to evaluate role of antioxidant (vitamin C) on modulation biochemical histological, histochemical and ultrastructural changes of liver caused by chronic use of monosodium glutamate.

Material and methods

Sixty adult albino rats, 8-10 weeks of age, weighing 120-140 g. They were maintained on a standard laboratory conditions. Rats were divided equally into three groups. First group "control" (n= 20) received 1ml of physiological saline (0.9%) daily by oral route "orogastric tube" for three months. Second group (n= 20) received monosodium glutamate (6mg/gm of body weight/day, dissolved in 1 ml of physiological saline "0.9%", orally "orogastric tube") for three months also. Third group received monosodium glutamate (6mg/gm of body weight/day, dissolved in 1 ml of physiological saline "0.9%") and vitamin C (500 mg/kg of body weight of /day, orally "orogastric tube") for three months .

Monosodium glutamate was obtained from Morgan Company for Chemical Industry (Tenth of Ramadan City- Egypt). Vitamin C (L- ascorbic acid) was obtained from Egyptian pharmaceutical industries company.

Blood samples were collected from hearts of rats of all groups on the last day (the ninety day). These samples centrifuged at 3000 rounds for 10 minutes for separation of the serum. This serum was analyzed for the following liver functions tests:

1- Aspartate Aminotransferase "AST"

AST is increased in the serum after hepatic tissue necrosis because most of the circulating AST activity is derived from the hepatocytes isoenzyme which presents in the liver cell (Crawford, 2006). Assay of AST was performed by mixing the serum to buffered solution of L-aspartic acid and 2-ketoglutarate and then incubated for one hour at 37 °C. After incubation, 1 mm of DNPH and 0.4m of NaOH was added (Chatterjee, 1993).

2- Alanine Aminotransferase "ALT"

ALT is localized to hepatocytes alone with high specificity. It is increased after hepatic cellular injury (Sherlock, 1997). Assay of ALT was performed by mixing the serum to buffered solution of DL- alanine and 2- ketoglutarate, and then incubated for thirty minutes at

37°C. After incubation, 1 mm of DNPH and 0.4m of NaOH was added (Daniel and Marshall, 1999).

3- Alkaline Phosphatase "ALP"

Elevation of serum alkaline phosphatase is resulted from any disease which will affect hepatocyte secretion because it is represented near the canalicular membrane of the hepatocyte (Pincus and Abraham, 2006). Assay of ALP was performed by using p-nitrophenol phosphate as substrate, in alkaline buffer with fresh unhemolysed serum for 45 min at 12°C (Daniel and Marshall, 1999).

Histological studies using light microscope

All animals were sacrificed after three months. Abdominal viscera were exposed by midline incision. The liver from the three groups were rapidly excised, cut into small pieces and dropped in formalin in which they were kept for appropriate time. After fixation, they were subjected to the normal procedure for paraffin embedding. Sections were cut at the thickness of 5 microns and stained with Haematoxyline-Eosin and Periodic acid Schiff reaction (PAS) and then tissue sections were investigated using light microscope (Drury and Wallington, 1980).

Histological studies using transmission electron microscope

Small pieces (1mm) of treated tissues were cut and fixed in 3% glutaraldehyde (pH 7.4) in phosphate buffer and post fixed in 2% osmium tetroxide in phosphate buffer. Following fixation, tissues were dehydrated at increasing concentrations of ethanol. They were then embedded in araldite resin. Ultrathin sections were cut using an ultratome. Ultrathin sections were stained by uranyl acetate saturated in 70% ethanol and lead citrate. Tissue sections were evaluated using a JEOL transmission electron microscope JEM-1200. Ex, Japan (Bancroft and Gamble, 2002).

Immunohistochemical studies

Immunohistochemistry is a localization of antigens or proteins in tissue sections by the use of labeled antibodies as specific reagents through antigen-antibody interactions that are visualized by a marker such as fluorescent dye or enzyme. Immunohistochemical studies was done and staining was performed using Immunohistochemistry stainer (DAKO Auto stainer Link48 _ Serial NO: As1217D0904) a standard avidin-biotin peroxidase complex (ABC) method (Heras et al., 1995).

The Ki-67

Antigen Ki-67 is a nuclear protein which is necessary for cellular proliferation and associated with ribosomal RNA transcription. Inactivation of this antigen leads to inhibition of ribosomal RNA formation. Positive expression of this antigen (Ki-67) indicates to the

increasing of cellular proliferation (Rahmanzadeh et al., 2007).

The p53

It is a tumor suppressor protein in the human and is encoded by the TP53 gene. It has an important role in apoptosis (cell death), genomic stability, and inhibition of angiogenesis. Positive expression of this protein (p53) indicates to tumor suppressor gene mutation (Kametaka et al., 2002).

The procedure

The Ki-67 and p53 proteins expression were examined using Ki-67 and p53 antibodies with dilution 1: 200 and 3µg/ml for 30 minutes at room temperature respectively (Lab Vision Corporation, USA). With reference to the manufacturer's specifications, these antibodies reacts broadly with all known Ki-67 and P53 variants of human, rat and mouse origin by immunohistochemistry. Each set of experiments included a positive control and ensured the reproducibility of the staining process. A negative control, in which the primary antibody was omitted, was also included in each set of experiments. The positive localization result was nuclear staining in two markers (Ki-67 and P53). Each section was counted manually at high power (X400) after identifying at low power (x100) the representative areas with the highest concentration of stained cells according to the recommendation of (Andrew et al., 2010). About 1000 cells/slide were counted in each of five microscopic fields from well-labeled areas to determine the average of Ki-67 and P53 labeling index (LI). LI was expressed as number of labeled cells (positive for stain) as a percentage of the total number of cells counted in each specimen. All identifiable staining was regarded as positive.

Statistical analysis

Results of this study were analyzed by using non - parametric test (Mann Whitney U test), Chi-Square test and independent sample T-test for comparisons.

Ethical considerations

The most appropriate animal species was chosen for this research. Promotion of a high standard of care and animal well-being at all times was done. Appropriate sample size was calculated by using the fewest number of animals to obtain statistically valid results. Surgical or other painful procedures were performed with appropriate sedation to avoid distress and pain. Our standards of animal care and administration met those required by applicable international laws and regulations.

Results

Biochemical changes

Table (1) shows mean \pm SD values of liver enzymes of rats (ALT, AST, and ALP). Mean \pm SD values of ALT were 11.25 ± 4.30 in control group which received saline 0.9%, 48.9 ± 14.3 in second group which received

monosodium glutamate dissolved in saline 0.9% and 24.3 ± 2.20 in third group which received monosodium glutamate dissolved in saline 0.9% and vitamin C. Mean \pm SD values of AST were 31.66 ± 3.70 in control group which received saline 0.9%, 53.25 ± 9.3 in second group which received monosodium glutamate dissolved in saline 0.9% and 36.13 ± 1.91 in third group which received monosodium glutamate dissolved in saline 0.9% and vitamin C. Mean \pm SD values of ALP were 88.77 ± 6.4 in control group which received saline 0.9%, 132.35 ± 10.2 in second group which received monosodium glutamate dissolved in saline 0.9% and 92.12 ± 3.3 in third group which received monosodium glutamate dissolved in saline 0.9% and vitamin C.

The previous data of table (1) showed that administration of MSG alone in the second group produced a statistical significant elevation of liver enzymes values when compared with the control group. Administration of vitamin C with MSG in third group leads to statistical significant difference of liver enzymes values when compared with the second group.

Macroscopic appearance

Liver rats of second and third groups were pale color with congestion and edema. Hepatic borders were lost. When the liver was cut, there were signs of edema and congestion.

Histopathological findings by light microscope

Examination of the liver control rats showed normal hepatic structure (Fig.1&2). But liver sections of second group rats showed loss of its characteristic architecture compared with the control group, the cytoplasm of the hepatocytes was characterized by coarse, pink, increased number of vacuoles and inflammatory cellular infiltration was abundant around the central vein with small fragmented pyknotic nuclei (Fig. 3). With more dilatation of central vein and blood sinusoids with degenerated vacuolated cytoplasm and multiple pyknotic nuclei (Fig.4). Liver sections of third group rats showed hypertrophy of hepatocytes and mild hypertrophy of kupffer cells with marked diminution of hydropic degeneration (Fig.5&6).

Histochemical results

1- Glycogen content

Control liver sections stained with periodic acid schiff's (PAS) showed mucopolysaccharide granules in the cytoplasm of hepatocytes, but the peripheral zonal cells showed higher mucopolysaccharide content than the central zonal cells (Fig.7). Liver sections of second group showed severe reduction of total amount of PAS positive material (Fig.8). Liver sections of third group showed mild reduction of PAS positive material but not reached to control group (Fig.9).

2- Collagen content

Control liver sections showed normal distribution of collagen and small amount of wavy fibrils (Fig.10). Collagen wavy fibrils either singly or fused together in dense bundles was observed in the second group (Fig.11). Liver sections of third group showed improvement in collagen deposition and connective tissue fibers as compared to liver of second group rats (Fig. 12).

Histopathological findings by transmission electron microscope

The ultrastructure of liver control rats showed that the hepatocytes were separated by blood sinusoid which lined by endothelial cells. The cytoplasm has a finely granular appearance due to the presence of numerous free glycogen granules. The mitochondria were numerous, usually round to oval in shape. The nuclei of the hepatocytes were oval or rounded with nucleoplasm consisting of a finely granular component, thin peripheral heterochromatin and euchromatin condensation. The sinusoidal lumina were lined by flat endothelial cells or their extensions which were separated from the adjacent hepatocytes by the subendothelial space or space of Disse (Fig.13&14).

The hepatocytes of liver of second group showed vacuolated cytoplasm (V), swollen mitochondria (M) and vesiculated rough endoplasmic reticulum (VER) with picknotic nuclei (Fig.15); dense collection of inflammatory cells including macrophages was observed in the cytoplasm (M) and the nuclear membrane is corrugated (Fig.16). Vacuolation (V) and dilatation of endoplasmic reticulum with no obvious ribosomes (RER) and atrophied mitochondria with ill- differentiated cisternae were noted (Fig.17).

The ultrastructural changes of the hepatocytes of liver of third group showed improvement in the form of diminution of cytoplasmic dissolution, well-defined nucleolus and nuclear envelope. Moreover, there were normal-shaped rough endoplasmic reticulum, but mitochondria were still dilated (Fig.18).

Table (2) shows P Values of histopathological results comparing between different groups using Chi-Square Test. P value of histopathological examination by light microscope of the second group which received MSG in comparison with first group which received normal saline (control) was 0.02. P value of histopathological examination by electron microscope of the second group which received MSG in comparison with the first group which received normal saline (control) was <0.01. P value of histopathological examination by light microscope of the third group which received MSG with vitamin C in comparison with the first group which received normal saline (control) was 0.01. P value of histopathological examination by electron microscope of the third group which received MSG with vitamin C in comparison with the first group

which received normal saline (control) was 0.02. P value of histopathological examination by light microscope of the third group which received MSG with vitamin C in comparison with the second group which received MSG was 0.04. P value of histopathological examination by electron microscope of the third group which received MSG with vitamin C in comparison with the second group which received MSG was < 0.01.

The previous data of table (2) showed statistically significant difference of histopathological results using the light and electron microscope in second group that received MSG alone in comparison with the control group. There was statistically significant difference of histopathological results using the light and electron microscope in third group which received vitamin C and MSG in comparison with the second group which received MSG alone. There was also statistically significant difference of histopathological results using the light and electron microscope in third group which received vitamin C and MSG in comparison with the control group.

Immunohistochemical results

The immunohistochemical study showed great differences in positivity of staining among different groups. The liver sections of control group rats which received saline and third group rats which received MSG and Vitamin C stained with Ki-67 showed slightly faint staining nuclei indicating the mild cell division of some hepatocytes (Fig.19). However, there was a strong immunoreactivity in liver section of second group rats which received MSG (Fig.20).

Table (3) explains the changes in liver Ki-67 and p53 indices. Rats of second group which received MSG were highly significant, while rats of third group which received MSG and vitamin C were high significant increase in compared with control rats and high significant when compared with second group (Fig.21). On the other hand, we used P53 as the Pro-apoptotic marker, where the liver sections of control and second groups were significant expression of p53. However, the liver sections of third group showed very weak positive stained nuclei (less than 10% of total cells) (Fig. 22, 23, 24).

The previous data of table (3) showed that administration of MSG alone in the second group produced a statistical significant difference of hepatic Ki-67 and P53 values when compared with the control group. Administration of vitamin C with MSG in third group leads to statistical significant difference of hepatic Ki-67 and P53 values when compared with the second group which received MSG alone.

Table (1): Effect of chronic use of monosodium glutamate on mean + SD of liver enzymes of rats.

Enzymes Group	ALT (IU/L)	AST (IU/L)	ALP (IU/L)
Group 1	11.25±4.30	31.66±3.70	88.77±6.4
Group 2	48.9±14.3*	53.25±9.3*	132.35±10.2*
Group 3	24.3±2.20**	36.13±1.91**	92.12±3.3**

ALT= alanine aminotransferase; AST= aspartate aminotransferase; ALP= alkaline phosphatase.

Group 1= Control (received saline 0.9%); Group 2= received monosodium glutamate dissolved in saline 0.9%; Group 3= received monosodium glutamate dissolved in saline 0.9% and vitamin C.

* = $p < 0.05$ (significant difference in comparison with control group); ** = $p < 0.05$ (significant difference in comparison with second group). Statistical analysis was performed by non - parametric test (Mann Whitney U test).

Table (2) P values of histopathological results comparison between different groups using chi-square test.

Pathological exam Group	Light Microscope	Electron Microscope
G ₂ Vs G ₁ *	0.02	<0.01
G ₃ Vs G ₁ **	0.01	0.02
G ₃ Vs G ₂ ***	0.04	< 0.01

$P < 0.05$ is statistically significant.

*Comparison between G₂ (received MSG dissolved in saline 0.9%) and G₁ (Control which received saline 0.9%);

Comparison between G₃ (received MSG dissolved in saline 0.9% and vitamin C) and G₁ (Control which received saline 0.9%); *Comparison between G₃ (received MSG dissolved in saline 0.9% and vitamin C) and G₂ (received MSG dissolved in saline 0.9%) .

Table (3): Comparison of hepatic Ki-67 and P53 indices among different groups.

Group Antibody	Control		Second		Third	
	Ki-67	P53	Ki-67	P53	Ki-67	P53
X ⁻	9.832	4.210	81.734	38.841	7.351	5.232
SD	5.811	0.913	11.981	8.221	5.132	1.231
SE	0.771	0.240	3.701	2.732	0.577	0.133
T-test	-	0.720	16.971	10.024	2.011	1.223
P value	-	-	≤0.01	≤0.006	≤0.03	≤0.001

Control group = first group, which received saline 0.9%; the second group received monosodium glutamate dissolved in saline 0.9%; the third group received monosodium glutamate dissolved in saline 0.9% and vitamin C.

X⁻= mean value; SD = standard deviation; SE = standard error

Statistical analysis was performed by t-test (non-parametric test); $P < 0.05$ is statistically significant.

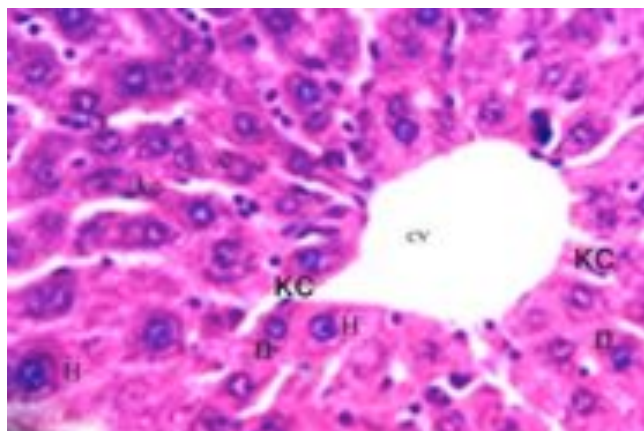


Fig. (1) Shows normal histological structure of liver of control rats. H&E (x400)

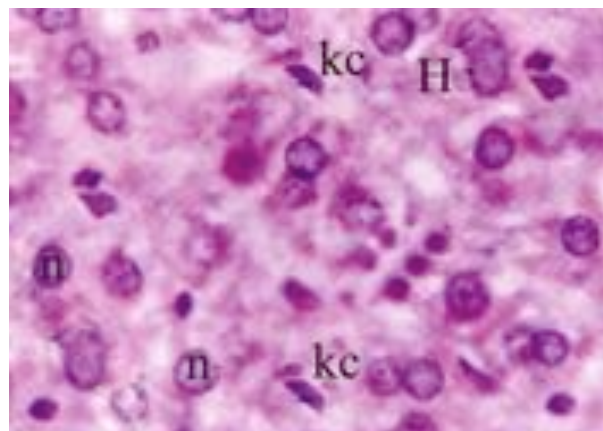


Fig. (2) Shows normal histological structure of liver of control rats. H&E (x1000)

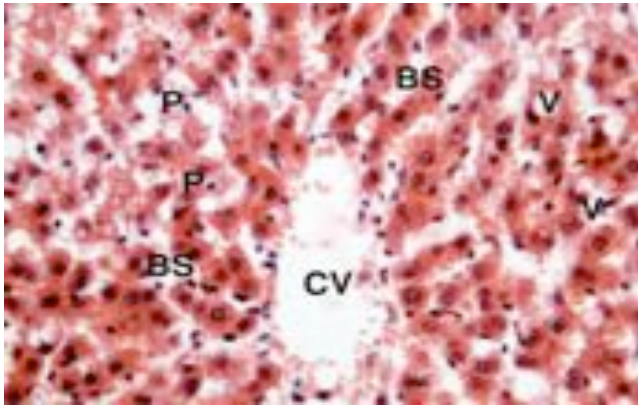


Fig. (3): Transverse section of liver rat of second group, shows dilatation of central vein (CV) and blood sinusoids (BS) with degenerated vacuolated (V) cytoplasm with multiple pyknotic nuclei (P). H&E (x400)

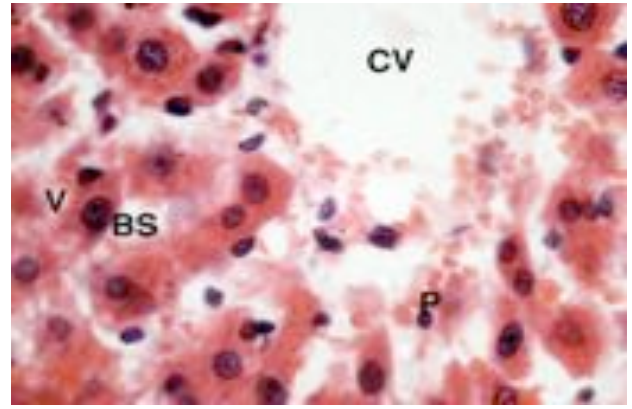


Fig. (4): Transverse section of liver rat of second group, shows dilatation of central vein (CV) and blood sinusoids (BS) with degenerated vacuolated (V) cytoplasm with multiple pyknotic nuclei (P). H&E (x1000)

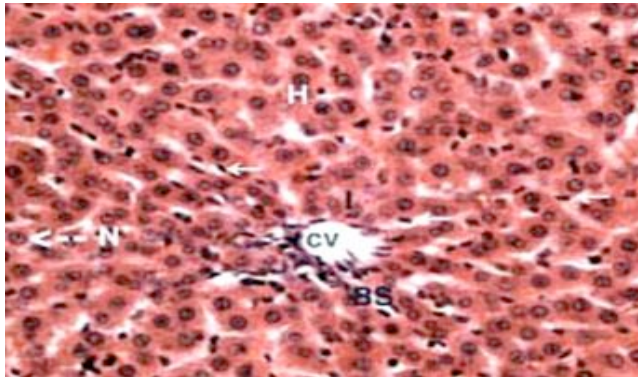


Fig. (5): Section of liver rat of third group shows normal central vein (CV) and hepatocytes (H) with normal nuclei (dotted arrow N) and inflammatory cell infiltration (I) with increasing in Kupffer cell (arrow). H&E (x400)

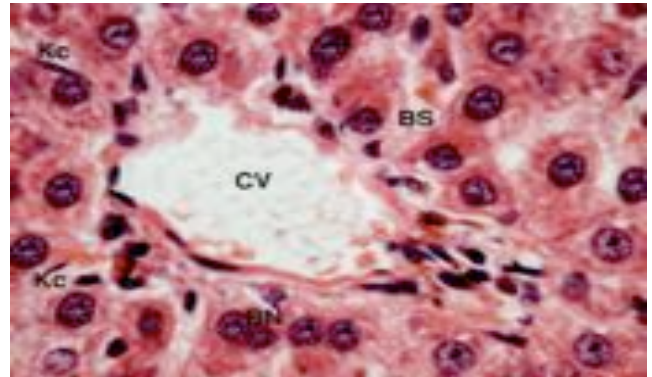


Fig. (6): Section of liver rat of third group shows hepatocytes with vacuolated cytoplasm (V) and separated by blood sinusoids (BS) which contain Kupffer cells (kc) and some binucleated (BN) cells. H&E (x1000)

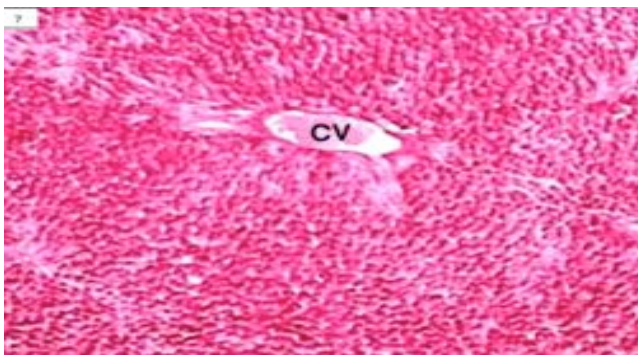


Fig. (7): Section of liver rat of control group shows PAS positive material in the cytoplasm of hepatocytes and normal central vein (CV), but nuclei has not any stains. PAS reaction (x200)

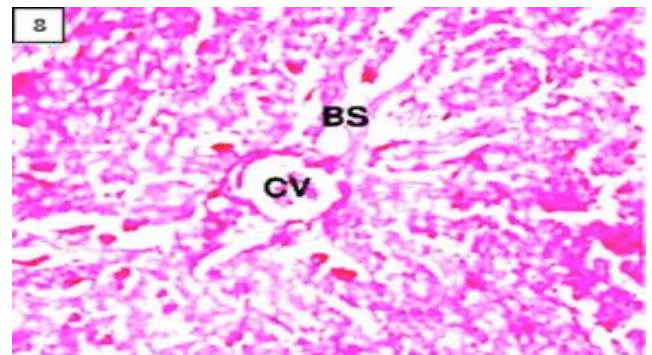


Fig. (8): Section of liver rat of second group shows decreased of PAS positive material in the cytoplasm (V) of hepatocytes, central vein (CV) blood sinusoids (BS). PAS reaction (x400)

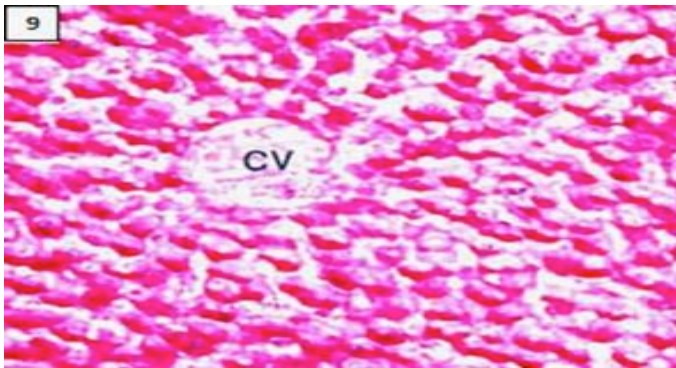


Fig. (9): Section of liver rat of third group shows improvement of PAS positive of cytoplasm of hepatocytes. PAS reaction (x400)

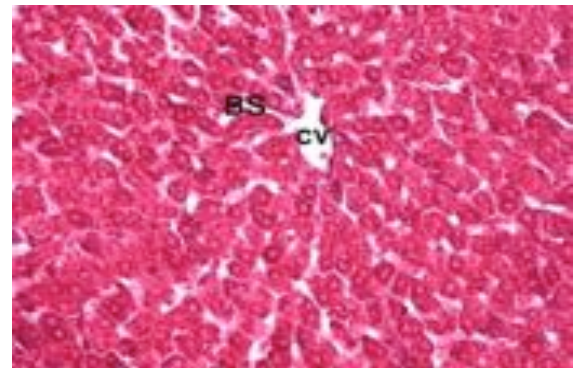


Fig. (10): Section of liver rat of control group shows normal distribution of collagen, with normal central vein (CV) and blood sinusoids (BS). Mallory stain (x200)

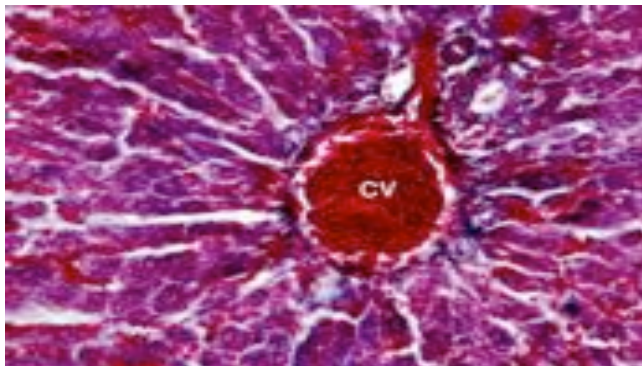


Fig. (11): Section of liver rat of second group shows increasing in collagen fiber especially around the central vein (CV) and in between hepatocytes. Mallory stain (x400)

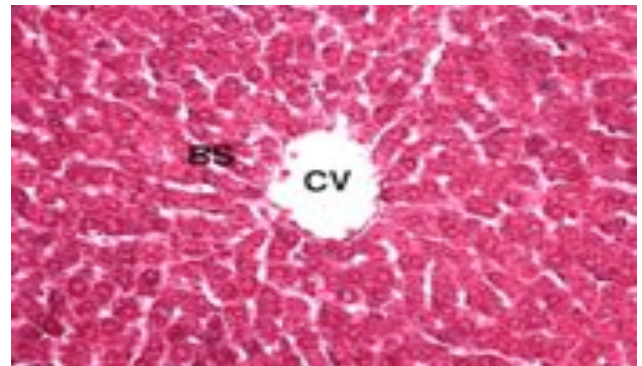


Fig. (12): Section of liver rat of third group shows normal distribution of collagen with normal central vein (CV) and blood sinusoids (BS). Mallory stain (x400)



Fig. (13): Liver of control rats shows rounded nucleus (N) of hepatocytes with slightly condensed peripheral heterochromatin (HE) and euchromatin (EU), rounded mitochondria (M). Uranyl acetate & lead citrate (x12000)

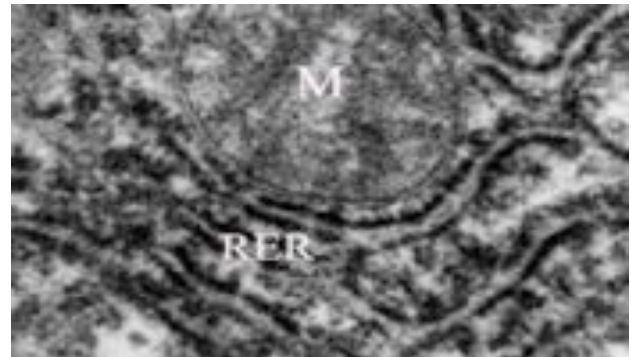


Fig. (14): Liver of control rats shows cytoplasm of hepatocytes which is crowded with organelles, particularly rough endoplasmic reticulum (RER) and normal mitochondria (M). Uranyl acetate & lead citrate (x18000)

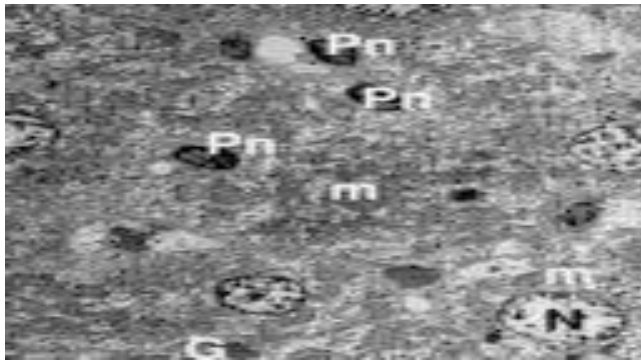


Fig. (15) : Liver of second group shows many hepatocytes which has signs of picknosis as irregular small nuclei (Pn) with many swollen mitochondria (m) and some cells appear normal nuclei (N) and many glycogen granules (G). Uranyl acetate & lead citrate (x2500)

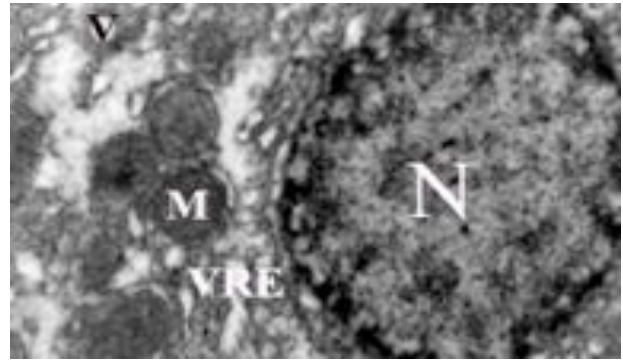


Fig. (16): Liver of second group shows hepatocytes with vacuolated cytoplasm (V), swollen mitochondria (M) and vesiculated rough endoplasmic reticulum (VER) with picknotic nucleus (N). Uranyl acetate & lead citrate (x8000)

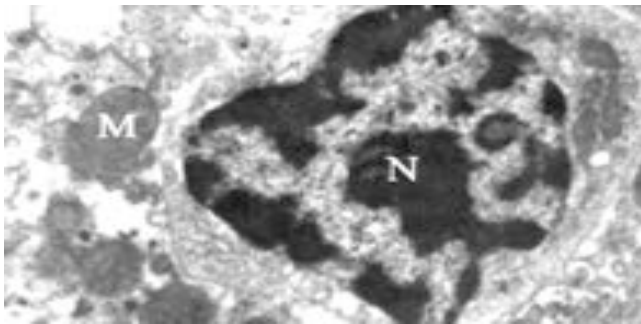


Fig. (17): Liver of second group shows hepatocytes with dense collection of inflammatory cells including macrophages (M) and corrugated nuclear membrane (N). Uranyl acetate & lead citrate (x10000)



Fig. (18): Liver of third group shows hepatocytes with large rounded euchromatic nucleus (N), well-defined nucleolus (n), nuclear envelope and dilated mitochondria (M). Uranyl acetate & lead citrate (x6000)

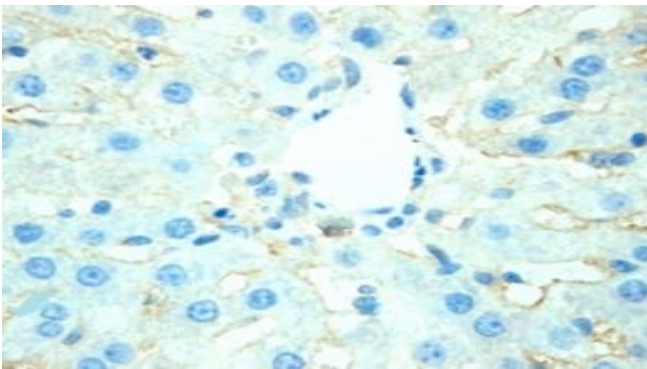


Fig. (19): Section of liver rat of control group shows negative staining of ki-67 in most of hepatocytes. Immunohistochemical staining (x400)

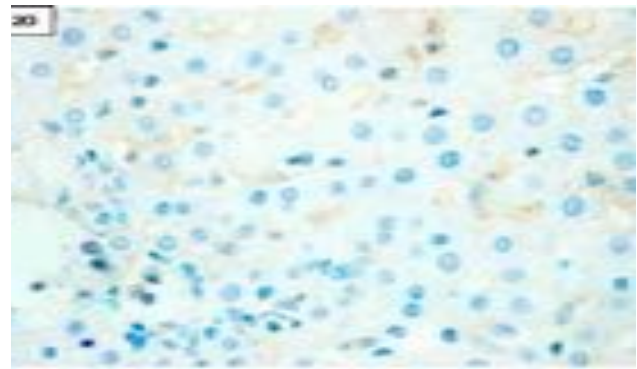


Fig. (20): Section of liver rat of control group shows negative staining of p53 in most of hepatocytes. Immunohistochemical staining (x400)

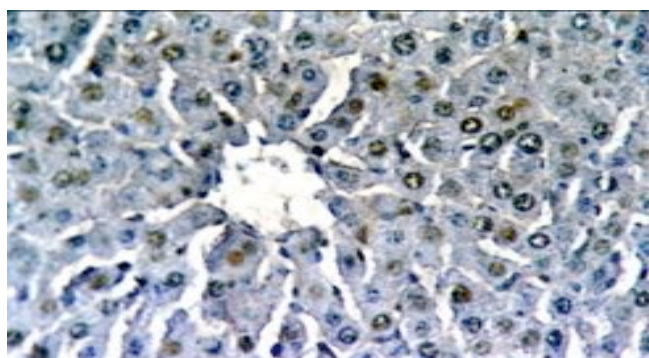


Fig. (21): Section of liver rat of second group shows strong staining of ki-67 in most of hepatocytes. (Immunohistochemical staining X 400)

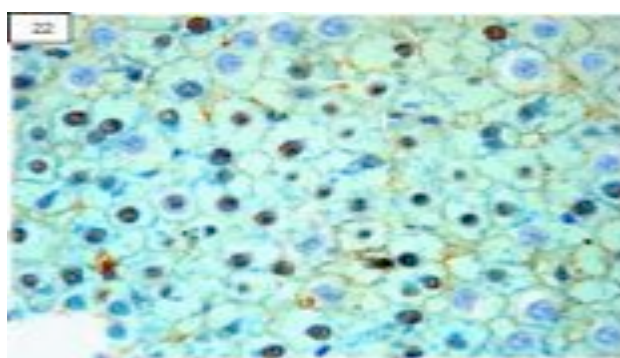


Fig. (22): Section of liver rat of second group shows strong staining of p53 in most of hepatocytes. (Immunohistochemical staining X 400)

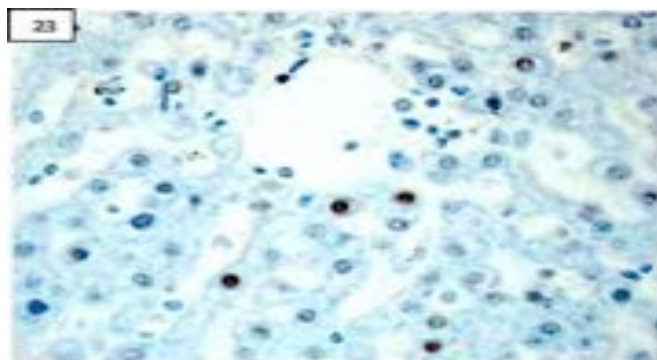


Fig. (23): Section of liver rat of third group shows faint immunoreactivity of ki-67 in nuclei of some hepatocytes. (Immunohistochemical staining X 400)

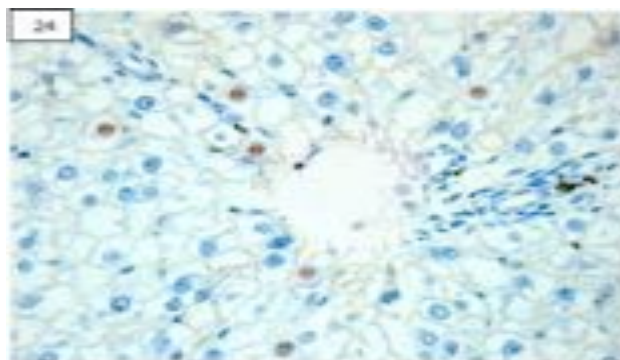


Fig. (24): Section of liver rat of third group shows faint immunoreactivity of p53 in nuclei of some hepatocytes. (Immunohistochemical staining X 400)

Discussion

Monosodium glutamate (MSG) is one of the most common food additives, which is widely used as a popular flavoring agent in canned foods. This study showed hepatotoxicity of monosodium glutamate and the effect of antioxidants agents such as vitamin C for modulation this toxicity. We performed this study by three groups of adult albino rats. The first group was control and second group received MSG alone while third group received MSG and vitamin C. We used the liver function tests to evaluate biochemical changes. Assessment of histopathological changes of liver was done by using light microscope, transmission electron microscope, histochemical studies, and immunohistochemical studies.

The present study showed statistical significant increase of liver enzymes activities values such as ALT, AST and ALP in second group which received monosodium glutamate alone in comparison with control group. Sherlock (1997) referred that the use of liver function tests is associated with high specificity especially when more than one test is abnormal.

AST is present in both the mitochondria and hepatocytes while ALT is localized to hepatocytes alone. The hepatocytes and mitochondrial forms of AST are true isoenzymes and immunologically distinct. About 80% of AST activity in liver is contributed by the

mitochondrial isoenzyme, whereas most of the circulating AST activity is derived from the hepatocytes isoenzyme. Large increases in mitochondrial AST occur in serum after extensive tissue necrosis. Mitochondrial AST is increased in chronic liver disease. Serum alkaline phosphatase is a heterogeneous group of enzymes. Hepatic alkaline phosphatase is represented near the canalicular membrane of the hepatocyte. Accordingly, diseases that predominately affect hepatocyte secretion will be accompanied by elevations of alkaline phosphatase levels (Pincus and Abraham, 2006)

AST and ALT were increased as result of hepatic necrosis which induced by chronic administration of MSG. The current study showed cellular hypertrophy and degenerative changes in the liver cell of second group which received monosodium glutamate as result of the cytotoxic effect of MSG on the liver, and then the detoxification processes and other functions of the liver will be affected and this consistent with Eweka et al., (2011). In third group which received monosodium glutamate and vitamin C, Liver function tests values (ALT, AST and ALP) were statistical significant decreased in comparison with second group which received monosodium glutamate alone. Generation of reactive oxygen species (ROS) as result of oxidative stress of MSG toxic effect were depressed by antioxidant

effect of vitamin C and this consistent with Oscar et al., (2006).

Sakr et al., (2003) suggested that toxic effect of MSG on the liver reflects on histopathological changes such as distortion of hepatocytes and dilatation of central vein of the liver. He reported that MSG administration increases the enzyme activity (ALT and AST) as result of hepatocellular injury and cause increased ratio of AST-to-ALT by 56.59% indicating possible cirrhosis and this consistent with Beyreuther et al., (2007) who showed that rat liver metabolize glutamate by oxidative deamination and then if large amounts of glutamate are ingested, hepatic metabolism of glutamate and toxic metabolites will be increased.

Our results showed that MSG caused foci of degeneration in the form of pyknosis of hepatocytes nuclei and fibrosis. These results were in agreement with Egbunu et al., (2009) who indicated that MSG induced hepatic necrosis and apoptosis, biliary proliferation, peribiliary fibrosis and degeneration of hepatocytes. Hazar et al., (2008) referred that oxidative stress which is responsible for the development of hepatic fibrosis and degeneration.

The present study showed that administration of MSG induced chronic inflammatory disease because of presence of inflammatory cells in the liver tissue and depletion of carbohydrates in the cytoplasm of hepatocytes. This consistent with Ortiz et al., (2006).

Our study showed that many ultrastructural abnormalities in the liver including lysosomes, endoplasmic reticulum, mitochondria and Golgi apparatus as result of cytotoxic effects of MSG. These results were in agreement with Iwase et al., (1998) who indicated also that Golgi apparatus is responsible for the packaging of hydrolytic enzymes involved in the formation of secretory products.

The current study showed that expansion of the mitochondrial intermembrane space and extension of the outer mitochondrial membrane leads to mitochondrial vacuolation and this consistent with Higgins et al., (2003) who referred also that vacuolation may be due to disturbance of ionic milieu of the cell with consequent retention of water and sodium leading to cellular swelling.

Our results showed that administration of vitamin C with administration of MSG leads to improvement in pathological changes of liver rats in comparison with group of rats which were administered MSG alone. These results confirmed that protection effect of vitamin C against toxicity of MSG because of its antioxidant action and free radical scavenger. These results consistent with Barros et al., (2004) and Pavlovic et al., (2009) who added also that vitamin C reverses some pathological changes which were induced in thymus of rats as result of administration of MSG. Results of Freeman (2006) explained that MSG is excitotoxin which causes hepatic damage by stimulating the generation of large numbers of free radicals and then,

modulation of hepatic toxicity of MSG was done by vitamin C.

Scholzen and Gerdes (2000) indicated that Ki-67 is a nuclear protein which is present in proliferating cells and expressed during late G1, S, M, and G2 phases of the cell cycle, while resting non-cycling cells (G0 phase) lacks of Ki-67 expression. Our results reported that high expression of ki-67 in the second group which was administered MSG only because it increases cell proliferation. In the third group which was administered vitamin C with MSG, there was the low expression of ki-67. So, vitamin C reduces cellular proliferation induced by MSG administration because it has an ability to regulate the expression and activity of cell cycle-related proteins and this consistent with Hahm et al., (2007).

The p53 antibody reacts with only mutant p53 protein and the half live of wild type is short "about 30 minutes" (Limaye et al., 2003). Our study revealed that presence the p53 over expression in the second group which received MSG alone. But, this over expression was decrease in control group and the third group which received vitamin C with MSG. So, our results confirmed that vitamin C may be caused alteration in tumor suppressor gene P53 by accumulate p53 protein because it inhibited this p53-induced Reactive Oxygen Species "ROS" generation and this consistent with Kim et al., (2008). Sung et al., (2011) indicated that another contrast fact which revealed that vitamin C induce P53 expression in colon cancer cell line.

Young and Woodside (2001) reported that Vitamin C is an antioxidant which removes free radicals products in the body and shows the ability to scavenge superoxide, hydrogen peroxide, and hydroxyl radicals. In the current study, the rats of control group and third group were similar in an immuohistochemical features, where the expression of both Ki-67 and P53 antibodies were weak positive in the previous two groups indicating that decrease of proliferation in case Ki-67 and mutation of P53 gene. So, our results revealed that the vitamin C has hepatoprotective effect on the parenchymal architecture of the liver against MSG because it caused significant decrease in proliferation cells and tumor suppressor genes mutation.

Conclusion

Chronic use of monosodium glutamate leads to hepatic toxicity which represents as biochemical, histopathological, and immunohistochemical changes in albino rats. Administration of Vitamin C as antioxidant agent with monosodium glutamate modulates hepatotoxicity induced by MSG.

Recommendations

Results of this study may be significant to animal but we need further researches in human to investigate our results. We suggest further studies with other parameters of liver assessment such as serum total protein, serum

albumin, serum total bilirubin, prothrombin time and determination of oxidative stress parameters such as extent of lipid peroxidation, reduced glutathione tissue level "GSH", superoxide dismutase "SOD" and catalase activity to complete this work. According to this study, chronic use of MSG leads to toxic effect in the liver and administration of vitamin C with MSG has a protective action against its hepatic toxicity.

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الملخص العربي

دراسة دور مضاد الأكسدة (فيتامين ج) في معادلة سمية الاستخدام المزمن لجلوتاميد أحادي الصوديوم في كبد الجرذان البيضاء

حسام الدين حسين عثمان¹ و سعيد سعيد الشماع² و أيمن المغاوري القناوى³

المقدمة: يستخدم جلوتاميد أحادي الصوديوم الآن في كثير من المواد الغذائية كإضافة للغذاء ومكسب للطعم. وبالرغم أنه يصنف كمكون غذائي آمن فإن استخدامه يبقى مثيراً للجدل. فجلوتاميد أحادي الصوديوم يعتبر من مضافات الأغذية ذات السم البطيء المثير والتي تسبب توالد كمية كبيرة من الشقائق الحرة والتي تؤثر على كثير من الأعضاء مثل الكبد. فيتامين ج يعمل كمضاد للأكسدة و يوقف عمل مسببات السرطان الكيماوية من خلال وقاية الجسم من الأكسدة. هدف الدراسة: ولذلك تهدف تلك الدراسة إلى دراسة دور مضاد الأكسدة (فيتامين ج) في معادلة التغيرات الكيماوية الحيوية و الهستولوجية و الهستوكيماوية والتركيبية الدقيقة في كبد الجرذان البيضاء الناتجة من الاستخدام المزمن لجلوتاميد أحادي الصوديوم.

المواد والطرق المستخدمة: ولقد أجري البحث على ستين من الجرذان البيضاء مقسمين إلى ثلاث مجموعات متساوية فالمجموعة الأولى الضابطة أخذت 1 مل من محلول الملح يوميا. والمجموعة الثانية أخذت 6 مل/جرام/يوميا من جلوتاميد أحادي الصوديوم. والمجموعة الثالثة أخذت 6 مل/جرام/يوميا من جلوتاميد أحادي الصوديوم و 500 مل/جرام/كجم يوميا من فيتامين سي بالفم و لمدة ثلاث شهور. تم فحص التغيرات الكيماوية الحيوية بعمل وظائف الكبد و التغيرات الهستوباثولوجية باستخدام الميكروسكوب الضوئي والالكتروني والفحوصات الهستوكيماوية والمناعية **النتائج:** بعد الاستخدام المزمن لجلوتاميد أحادي الصوديوم و بالفحص بالميكروسكوب الضوئي والالكتروني اكتشف تغيرات هستوباثولوجية مثل تدمير بنية الكبد و اتساع في الأوردة المركزية في الخلايا الكبدية مع زيادة ذات دلالة إحصائية في إنزيمات الكبد. كما أوضحت الفحوص الهستوكيماوية انخفاضا حادا في الكربوهيدرات والبروتينات و تغلظ في النواة وتكوين فجوات في السيتوبلازم وانتفاخ في الميتوكوندريا وتحوصل الشبكة الأندوبلازمية الخشنة مع وجود ايجابية ذات دلالة في صبغة ki-67 و p53 للنواة. ولقد ادي إعطاء فيتامين ج مع جلوتاميد أحادي الصوديوم إلى تحسن ذي دلالة في التغيرات الكيماوية الحيوية و الباثولوجية في الكبد.

الخلاصة: فذلك فان سمية كبد الجرذان الناتجة من الاستخدام المزمن لجلوتاميد أحادي الصوديوم تتحسن بإعطاء فيتامين ج معه

الكلمات المفتاحية: جلوتاميد أحادي الصوديوم فيتامين ج (ج) الكبد.

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